







# Predicting tropical tree mortality with leaf spectroscopy

Christopher E. Doughty<sup>1</sup>  | Alexander W. Cheesman<sup>2,3</sup> | Terhi Riutta<sup>4,5</sup> | Eleanor R. Thomson<sup>4</sup>  | Alexander Shenkin<sup>4</sup>  | Andrew T. Nottingham<sup>6,7</sup>  | Elizabeth M. Telford<sup>7</sup> | Walter Huaraca Huasco<sup>4</sup>  | Noreen Majalap<sup>8</sup> | Yit Arn Teh<sup>9</sup> | Patrick Meir<sup>10,7</sup> | Yadvinder Malhi<sup>4</sup> 

<sup>1</sup>School of Informatics, Computing, and Cyber Systems, Northern Arizona University, Flagstaff, AZ, USA

<sup>2</sup>College of Life and Environmental Sciences, University of Exeter, Exeter, UK

<sup>3</sup>College of Science & Engineering, James Cook University, Cairns, Qld, Australia

<sup>4</sup>Environmental Change Institute, School of Geography and the Environment, University of Oxford, Oxford, UK

<sup>5</sup>Department of Life Sciences, Imperial College London, Ascot, UK

<sup>6</sup>School of Geography, University of Leeds, Leeds, UK

<sup>7</sup>School of Geosciences, University of Edinburgh, Edinburgh, UK

<sup>8</sup>Forest Research Centre, Sabah Forestry Department, Sandakan, Malaysia

<sup>9</sup>School of Natural and Environmental Sciences, Newcastle University, Newcastle Upon Tyne, UK

<sup>10</sup>Research School of Biology, Australian National University, Canberra, ACT, Australia

## Correspondence

Christopher E. Doughty, School of Informatics, Computing, and Cyber Systems, Northern Arizona University, Flagstaff, AZ 86011, USA.

Email: [chris.doughty@nau.edu](mailto:chris.doughty@nau.edu)

## Funding information

John Fell Fund, University of Oxford; NASA Biodiversity, Grant/Award Number: 80NSSC19K0206; NERC, Grant/Award Number: NE/K01627X/1, NE/K016377/1; ERC, Grant/Award Number: GEM-TRAIT (321131); ARC grants, Grant/Award Number: DP17010409 and DP190101823

Associate Editor: Jennifer Powers

Handling Editor: Jennifer Powers

## Abstract

Do tropical trees close to death have a distinct change to their leaf spectral signature? Tree mortality rates have been increasing in tropical forests, reducing the global carbon sink. Upcoming hyperspectral satellites could be used to predict regions close to experiencing extensive tree mortality during periods of stress, such as drought. Here we show, for a tropical rainforest in Borneo, how imminent tropical tree mortality impacts leaf physiological traits and reflectance. We measured leaf reflectance (400–2500 nm), light-saturated photosynthesis ( $A_{\text{sat}}$ ), leaf dark respiration ( $R_{\text{dark}}$ ), leaf mass area (LMA), and % leaf water across five campaigns in a six-month period during which there were two causes of tree mortality: a major natural drought and a co-incident tree stem girdling treatment. We find that prior to mortality, there were significant ( $p < 0.05$ ) leaf spectral changes in the red (650–700 nm), the NIR (1,000–1,400 nm), and SWIR bands (2,000–2,400 nm) and significant reductions in the potential carbon balance of the leaves (increased  $R_{\text{dark}}$  and reduced  $A_{\text{sat}}$ ). We show that the partial least squares regression technique can predict mortality in tropical trees across different species and functional groups with medium precision but low accuracy ( $r^2$  of .65 and RMSE/mean of 0.58). However, most tree death in our study was due to girdling, which is not a natural form of death. More research is needed to determine if this spectroscopy technique can be applied to tropical forests in general.

## KEYWORDS

drought, el Niño, girdling, spectroscopy, traits, tree mortality, tropical forests

## 1 | INTRODUCTION

Can future tropical forest tree mortality be predicted with aircraft or satellite remote sensing? This question is of interest because tropical tree mortality is increasing in places, reducing the global carbon sink (Brienen et al., 2015; Hubau et al., 2020). Increased tree mortality may be driven by recent increases in extreme weather events caused by climate change, including increased drought frequency/severity (Doughty et al., 2015; Rifai et al., 2018, 2019; Rowland, Da Costa, et al., 2015) or elevated air temperatures (Clark, 2004; Doughty & Goulden, 2009a). Other causes of mortality include altered disturbance regimes due to land management practices or biological invasions (eg, grass/fire cycles) and the negative environmental impacts arising from forest degradation (eg, physical damage to trees from logging or small-scale slash-and-burn agriculture; environmental stress from enhanced edges effects) (Malhi et al., 2014). Experimental drought manipulations in the Amazon (da Costa et al., 2010; Meir et al., 2015; Nepstad et al., 2007) show that larger trees are more susceptible to drought-related mortality for specific high-abundance taxa (Bittencourt et al., 2020).

Could changes to leaf properties in these large trees indicate risk of imminent future mortality? Death of these large individuals has the greatest impact on tropical forest vegetation and carbon dynamics (Phillips et al., 2009). “Environmental surveillance” techniques that enable us to identify individuals at risk of mortality or to predict future patterns of senescence would enable us not only to model forest vegetation and carbon dynamics more accurately, but could possibly enable us to manage the spread of forest pathogens and understand environmental stress gradients related to disturbance. Given that these large trees are also the most visible to aircraft and satellites, remote sensing techniques that enable us to identify dying trees hold tremendous potential for detecting and understanding the causes of tree mortality at large spatial scales.

Leaf traits, such as leaf chemical composition, photosynthetic capacity or leaf mass per area (LMA), are important indicators of a tree's life history strategy and overall vitality (Poorter et al., 2008; Wright et al., 2004, 2010). Remote sensing of these traits is thus one approach that could enable us to detect individuals or taxa at elevated risk of death from stress. For instance, light-demanding species with rapid growth and high mortality rates are predicted to have lower seed mass, leaf mass per area (LMA), wood density, and tree height (Wright et al., 2010). Variation in LMA in part expresses a trade-off between the energetic cost of leaf construction and the light captured per area that may be reflective of the strategy of the broader tree itself (Díaz et al., 2016; Poorter et al., 2009). Drought tolerance is also reflected in structural traits such as LMA, leaf thickness, leaf toughness, and wood density, although further studies are required to better establish the limitations of these metrics and identify other potential indices (Bartlett et al., 2012; Fyllas et al., 2012; Niinemets, 2001; Zanne et al., 2010).

Much recent literature has discussed the roles of carbon starvation, hydraulic failure, or a combination of the two on tree death as well as the traits associated with these processes. To predict tree

death with remote sensing we must first understand the characteristics that drive tree death. A recent meta-analysis suggests that metrics of hydraulic failure more consistently predicted mortality than carbon starvation as determined by tissue concentrations of non-structural carbohydrates (NSC) (Adams et al., 2017). Another study similarly found hydraulic traits were better at predicting the response of ecosystem fluxes ( $\text{CO}_2$  and water vapor) to drought than traits like LMA or wood density (Anderegg et al., 2018). Tree mortality during droughts is highest for species that have a small hydraulic safety margin (the difference between typical minimum xylem water potential experienced and xylem vulnerability to embolism) (Anderegg et al., 2016). Turgor loss point—the leaf water potential that induces wilting—may be a key trait predicting drought tolerance and species distributions relative to water supply (Bartlett et al., 2012). In tropical forests, turgor loss point varied widely across species and was weakly positively correlated with leaf toughness and thickness (Maréchaux et al., 2015). Some literature suggests that both hydraulic failure and carbon depletion are associated with mortality in large part through their effect on leaf water content and turgor (Sapes et al., 2019; Sevanto et al., 2014). Leaf water content can be accurately remotely sensed at the leaf and aircraft scale (Asner et al., 2016; Asner & Martin, 2008).

Leaf traits can be sensed remotely by aircraft or from space. Foliar traits such as nitrogen (N) content, chlorophyll content, carotenoids, lignin, cellulose, LMA, soluble carbon, and water can be remotely estimated with leaf spectral reflectance signatures (400–2,500 nm in 1 nm bandwidths) in many different plants and ecosystems (Ustin et al., 2009), including tropical forests (Asner & Martin, 2008). This is because certain traits are associated with reflectance characteristics within specific spectral regions. For instance, the visible part of the spectrum (400–700 nm) is associated with pigments (mostly chlorophyll), and the near infrared (NIR; 700–1,300 nm) is associated with structures such as palisade cell density. LMA and leaf chemistry have been accurately measured and modeled at both the leaf (Asner & Martin, 2008; Curran, 1989; Jacquemoud et al., 2009), canopy and landscape scales (Asner et al., 2016). Other elements not directly expressed in the spectrum, such as phosphorus (P), have been accurately predicted with spectroscopy, possibly through stoichiometric relationships with other chemical species (Ustin et al., 2006, 2009) or correlations with leaf morphological traits via the leaf economics spectrum (Wright et al., 2004). Other tropical tree traits not directly associated with leaf spectra, such as photosynthetic capacity (Doughty et al., 2011), and branch wood density, have been predicted with spectroscopy in tropical forests (Doughty et al., 2017). Traits not directly associated with spectral regions can still be predicted through correlations between leaf traits and a tree's life history strategy (Doughty et al., 2017).

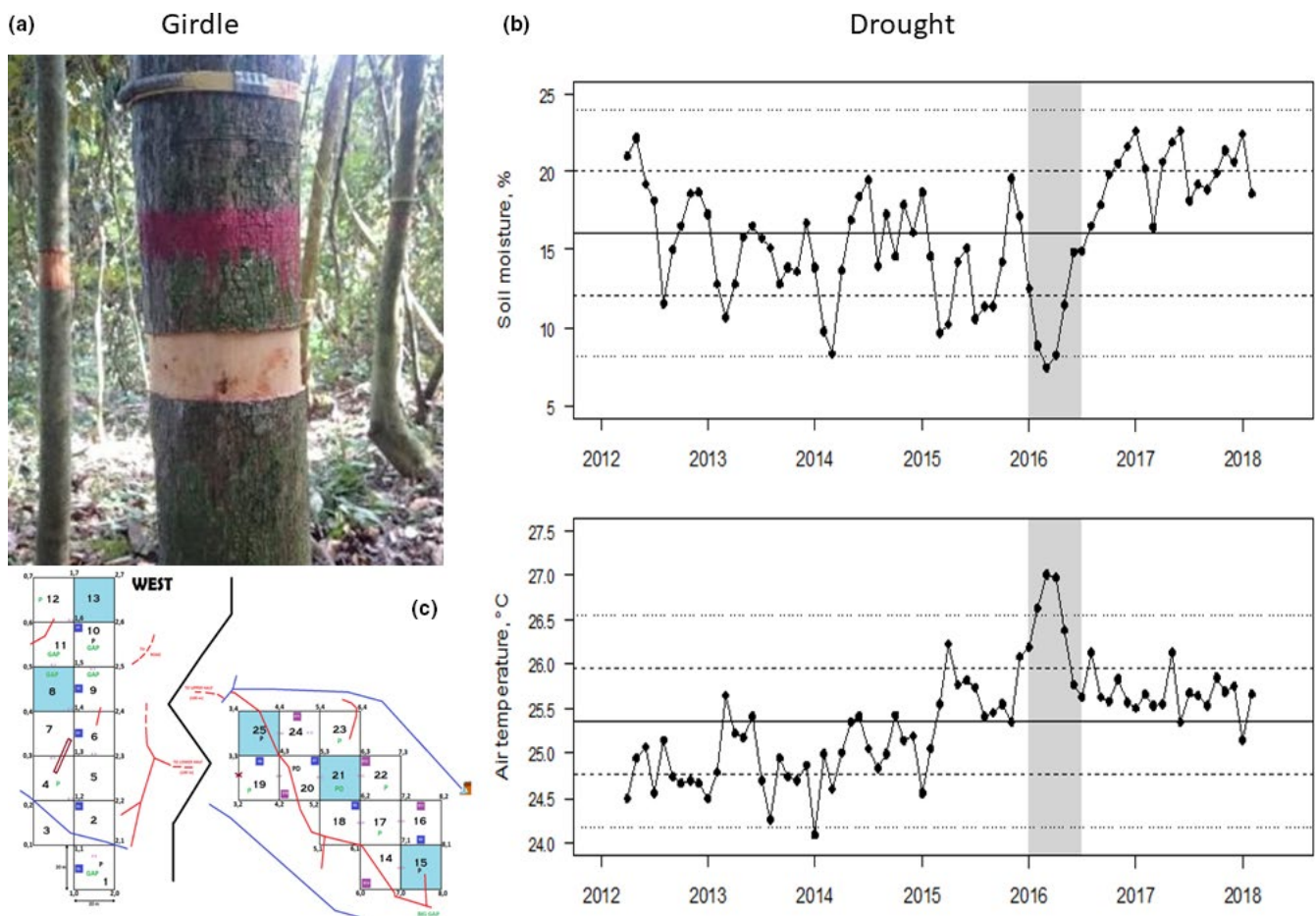
There is evidence that drought changes tropical forest reflectance at the continental scale, due to changes in leaf traits or increased tree mortality. For instance, Enhanced Vegetation Index (EVI), a greenness index measured with Moderate Resolution Imaging Spectroradiometer (MODIS), increased in the Amazon

during the 2005 drought, indicating possible positive impacts of drought on forests due to increased irradiance (Saleska et al., 2007). However, others have challenged the original interpretation of the EVI data (Morton et al., 2014; Samanta et al., 2010), highlighting the challenge of remote sensing at a continental scale. More recently, during a major El Niño drought in Borneo, NDVI initially increased as the drought was strengthening, but decreased at its peak (Nunes et al., 2019). Interpretation of changing NDVI and/or EVI at larger spatial scales is generally complicated in many ecosystems as changes at the leaf level may be compensated for or masked by canopy scale process. For example, leaf senescence and leaf fall may reduce the canopy scale NIR signal. However, remotely sensed LAI signal saturates in tropical forests and LAI variation can be relatively small even following strong climate extremes such as drought. For instance, Meir et al. (2018) found a 12%–20% change in LAI during an extreme drought manipulation experiment with a non-droughted natural LAI of  $\sim 5.5 \text{ m}^2 \text{ m}^{-2}$ , which is within the saturation range. Therefore, changes in tropical forest canopy spectral characteristics at larger spatial scales may be more linked to changes in leaf-level

spectra, than in other ecosystems with lower LAI (Doughty & Goulden, 2009b; Wu et al., 2018).

The 2016 El Niño caused a significant drought in Borneo, both in terms of increased maximum temperatures and reduced precipitation (Figure 1) (Rifai et al., 2019; Rifai et al., 2018). This El Niño had unusually high temperatures, which have been attributed to climate change (Thirumalai et al., 2017). Recent work in Borneo, near our study site, found the El Niño event was associated with a decrease in chlorophyll and carotenoid concentrations by 35%. They also noted a decrease in NDVI with a change in the shortwave infrared region of leaf spectral signatures (Nunes et al., 2019). The authors hypothesized that trees produced new leaves with higher pigment concentrations at the start of the El Niño event, and then dropped their leaves at its peak.

In this study, we focus on tree mortality at a 1 ha long-term study site close to the Nunes et al. (2019) study site in Sabah, northern Borneo. We attempt to understand the relationship between leaf traits, spectroscopy, and mortality in two different ways: natural death during El Niño and forced mortality induced by girdling.



**FIGURE 1** (a) An example tree that was girdled by stripping 10 cm of phloem in a ring around the tree. (b) Monthly volumetric soil moisture content at 20 cm depth (top) and air temperature (bottom) records at the study site. The horizontal continuous line denotes the long-term mean and the dashed lines denote 1 and 2 standard deviations. The gray region is the period of our measurements. (c) A schematic of the plot layout with the non-girdled trees in the section labeled West (the other section was girdled). The total area of the plot is 1 ha, with the two sections separated by approximately 200 m. The middle black line represents the river. Each individual square represents a 20 m × 20 m subplot. Red lines are trails, and blue lines are small temporary streams

Before, during and after the 2016 El Niño drought (over 5 field campaigns), we measured canopy-top leaf spectra (400–2,500 nm), net light-saturated photosynthesis, dark respiration, % water and LMA in a representative cross-section of the 393 monitored trees. We further tried to explore mechanisms of mortality with a girdling campaign (the removal of the phloem in a 10 cm ring around the tree stem) in one half (0.5 ha, 210 stems) of the plot. Here, we test the following hypotheses:

H1 – Leaf traits that are correlated with leaf spectroscopy signals, such as light-saturated photosynthesis, dark respiration, % water and LMA, undergo significant change months prior to tree mortality.

H2–Tropical tree mortality can be predicted with hyperspectral information (400–2,500 at 1 nm band-width leaf reflectance).

## 2 | METHODS

### 2.1 | Study sites

Our study plots are in Kalabakan Forest Reserve in Sabah, Malaysian Borneo (Tower SAF-05 4.716°, 117.609°) within the Stability of Altered Forest Ecosystems (SAFE) Project study site (Ewers et al., 2011; Riutta et al., 2018). A schematic of the study site is shown in Figure 1c. Mean annual temperature is approximately 26.7°C, and mean annual precipitation is 2,600–2,700 mm with no distinct dry season but, on average, ~12% of months with precipitation below 100 mm/month (Walsh & Newbery, 1999). The plot has been selectively logged four times since the 1970s, which represents a high logging intensity for this region. The soils are orthic Acrisols or Ultisols on undulating clay soil. The tree basal area is 13.9 m<sup>2</sup>/ha. Total NPP and autotrophic respiration have been measured at this plot since 2011, and there is an eddy covariance tower nearby (Riutta et al., 2018). The plot is split in half by a small stream. All the trees on one side of the stream were girdled in late January 2016 by removing the phloem tissue in a 10 cm band, as described below (note: The plot was in the process of conversion to oil palm

agriculture production). This part of the study site is hereafter referred to as the “girdled plot.” The trees on the other side of the stream were not girdled and represent the treatment control. This part of the study site is hereafter referred to as the “drought plot.” Although all trees experienced drought, the “drought” plot only experienced drought and not the effects of girdling. We collected data during five field campaigns that took place from January to June 2016. Campaigns began on the following days and generally took several days: Campaign 1 = 21 Jan-16, Campaign 2 = 10 Feb-16, Campaign 3 = 01 Mar-16, Campaign 4 = 29 Mar-16, Campaign 5 08 Jun-16. The first field campaign (C1) was conducted before girdling occurred to determine pre-girdling conditions and process rates.

### 2.2 | Girdling experiment

In late Jan 2016, after the first field campaign, we further explored the causes of tree mortality by conducting a girdling experiment. Girdling involved removing a 10 cm strip of the periderm and phloem in a ring around the tree stem at ~ 1.2 m height (with exceptions for trees with buttresses, which were girdled above the buttress) above the soil (Figure 1a) in a plot that was scheduled for conversion to a palm oil plantation. This technique prevents carbohydrate transport to the roots but maintains hydraulic connectivity because xylem tissues are not severed. Tree death was determined visually, based on the absence of visible canopy, with regular (average 18-day period) visits to the plots for both the drought and the girdled plots. We give the species measured in both plots in Table 1.

### 2.3 | Leaf sampling strategy

In each plot, 20–25 trees were chosen during each campaign, and tree climbers with extendable tree pruners removed one branch per tree that was growing in full sunlight (Asner & Martin, 2008). These branches were quickly recut underwater and taken to the laboratory for further measurements. On each of these branches, five fully expanded non-senescent leaves in randomly selected locations were chosen for measurements of leaf gas exchange, leaf spectral properties (measured within 1 hr of being cut) and LMA. Leaf area was determined

**TABLE 1** Tree species measured intensively in the drought and girdled plot aligned to show which species were measured in both plots

Girdled Plot	Drought Plot
<i>Adinandra borneensis</i> , <i>Brownlowia peltata</i> , <i>Dryobalanops lanceolate</i> , <i>Duabanga moluccana</i> , <i>Hydnocarpus anomalus</i> , <i>Leea aculeate</i> , <i>Lithocarpus blumeanus</i> , <i>Litsea garciae</i> , <i>Lophopetalum</i> sp., <i>Macaranga hypoleuca</i> , <i>Macaranga pearsonii</i> , <i>Neolamarckia cadamba</i> , <i>Nephelium rambutan</i> , <i>Parashorea malaanonan</i> , <i>Shorea johorensis</i> , <i>Shorea parvifolia</i> .	<i>Adinandra borneensis</i> , <i>Cariumna odontophyllum</i> , <i>Diplodiscus paniculatus</i> , <i>Dipterocarpus caudiferus</i> , <i>Dryobalanops lanceolate</i> , <i>Duabanga moluccana</i> , <i>Endospermum peltatum</i> , <i>Lithocarpus blumeanus</i> , <i>Macaranga pearsonii</i> , <i>Mallotus leucodermis</i> , <i>Nauclea officinalis</i> , <i>Neolamarckia cadamba</i> , <i>Parashorea malaanonan</i> , <i>Pleiocarpidia sandakanica</i> , <i>Pterospermum elongatum</i> , <i>Shorea gibbosa</i> , <i>Shorea johorensis</i> , <i>Syzygium</i> sp., <i>Trema orientalis</i>

immediately after collection using a digital 476 scanner (Canon LiDE 110). Leaves were then oven dried at 72°C until constant mass was reached. We subtracted wet weight from dry weight to calculate % leaf water and used dry weight and leaf area in order to calculate LMA.

## 2.4 | Leaf-level gas exchange

We used a portable gas exchange system (LI 6400, Li-Cor Biosciences, Lincoln, NE, USA) to measure leaf-level gas exchange. After returning to the laboratory, leaf dark respiration ( $R_{\text{dark}}$ ) was measured by covering branches with an opaque bag for at least 20 min prior to measurement at a cuvette temperature of 30°C (Rowland et al., 2017). After this, branches were exposed to sunlight and light-saturated leaf photosynthesis was measured ( $A_{\text{sat}}$ ; 1,200  $\mu\text{mol}/\text{m}^2 \text{ s}^{-1}$  PPFD, 400 ppm  $\text{CO}_2$ , at 30°C). We chose a light level of 1,200  $\mu\text{mol}/\text{m}^2 \text{ s}^{-1}$  for  $A_{\text{sat}}$  because we tested photosynthetic capacity and found it generally saturated below light levels of  $\sim 1,200 \mu\text{mol}/\text{m}^2 \text{ s}^{-1}$  PPFD, similar to other tropical studies (Both et al., 2019; Doughty & Goulden, 2009b; Gvozdevaite et al., 2018). We waited for gas exchange values to stabilize before starting a measurement, recorded data every two seconds and averaged the results after eliminating the first 20 measurements. We excluded photosynthesis measurements less than 0  $\mu\text{mol}/\text{m}^2 \text{ s}^{-1}$  as this was indicative of a failure to maintain hydraulic connectivity in the sampled branch resulting in stomatal closure. We also excluded dark respiration measurements more negative than  $-1.5 \mu\text{mol}/\text{m}^2 \text{ s}^{-1}$  as this was considered indicative of a failure to truly represent  $R_{\text{d}}$  or in some cases operator error. Most physiological measurements were collected between 07:00 and 14:00 local time, and branches were cut from trees between 06:00 and 13:00 local time. Our data archive (Doughty et al., 2020) includes our averaged  $\pm$  sd data for each leaf measured for transpiration rate ( $\text{mmol H}_2\text{O m}^{-2} \text{ s}^{-1}$ ), vapor pressure deficit based on leaf temperature (kPa), intercellular  $\text{CO}_2$  concentration ( $\mu\text{mol CO}_2 \text{ mol}^{-1}$ ), conductance to  $\text{H}_2\text{O}$  ( $\text{mol H}_2\text{O m}^{-2} \text{ s}^{-1}$ ), and photosynthetic rate ( $\mu\text{mol CO}_2 \text{ m}^{-2} \text{ s}^{-1}$ ).

## 2.5 | Leaf spectroscopy

We randomly selected five leaves within an hour of each branch being cut and measured hemispherical reflectance near the mid-point between the main vein and the leaf edge (Asner & Martin, 2008). We used an ASD Fieldspec 4 with a fiber optic cable, contact probe and a leaf clip (Analytical Spectral Devices, Boulder, Colorado, USA). The spectrometer records 2,175 bands spanning the 325–2,500 nm wavelength region. We corrected for small discontinuities between spectral bands ( $\sim 950$  and  $\sim 1,750$  nm), where the instrument transitions from one sensor to another. Measurements were collected with 136-ms integration time per spectrum (Asner & Martin, 2008; Doughty, Asner, et al., 2011). To ensure measurement quality, the spectrometer was calibrated for dark current and stray light, and white-referenced to a calibration panel (Spectralon,

Labsphere, Durham, New Hampshire, USA) after each branch (Asner & Martin, 2008; Doughty et al., 2011). The spectrometer was optimized after every branch so the light levels did not saturate. For each measurement, 25 spectra were averaged together to increase the signal-to-noise ratio of the data.

## 2.6 | Data analysis

We used the partial least squares regression (PLSR) modeling approach to predict leaf traits with spectral information (Geladi & Kowalski, 1986). PLSR incorporates all the spectral information within each leaf reflectance measurement, eventually reducing all spectral data (400–2,500 nm) down to a relatively few, uncorrelated latent factors. This approach has been used successfully to predict plant traits across a wide range of ecosystems, including tropical forests (Asner & Martin, 2008; Serbin et al., 2014). We used the PLSregress command in Matlab (Matlab, MathWorks Inc., Natick, MA, USA) to establish predictive models for LMA,  $A_{\text{sat}}$ , wood density (estimated with tree species and a lookup table (Chave et al., 2009)) and tree mortality (Doughty, Asner, et al., 2011). To avoid over-fitting the number of latent factors, we minimized the mean square error with K-fold cross validation. To avoid issues of pseudo-replication, we emphasize that the unit of analysis in these analyses is the leaf. To create a completely independent testing dataset, we used the above method on 70% of our data to calibrate our model and then the remaining 30% to test the accuracy of our model. We evaluated the accuracy of our modeled estimates using two main metrics:  $r^2$  and root mean square error (RMSE)/mean. We graded our results as high precision and accuracy ( $r^2 > 0.70$ ; %RMSE < 15%), medium precision and accuracy ( $r^2 > 0.50$ ; % RMSE < 30%), low precision, and accuracy ( $r^2 > 0.50$ ; % RMSE > 30%). We also calculated NDVI for our five study periods as  $\text{NDVI} = (\text{NIR} - \text{red})/(\text{NIR} + \text{red})$  where we use 1,000 nm for NIR and 650 nm for red.

## 2.7 | Statistical tests

For our leaf spectral measurements, for each 1 nm bandwidth, we determined statistical significance ( $p < 0.05$ ) between trees within 50 days of mortality and prior to this with a paired  $t$  test (Matlab, Mathworks). To understand significant differences between % water, LMA,  $R_{\text{dark}}$ , and  $A_{\text{sat}}$ , we used a  $t$  test. To understand the impact of the girdling between % water, LMA,  $R_{\text{dark}}$ , and  $A_{\text{sat}}$  over time, we used a repeated measures ANOVA.

## 3 | RESULTS

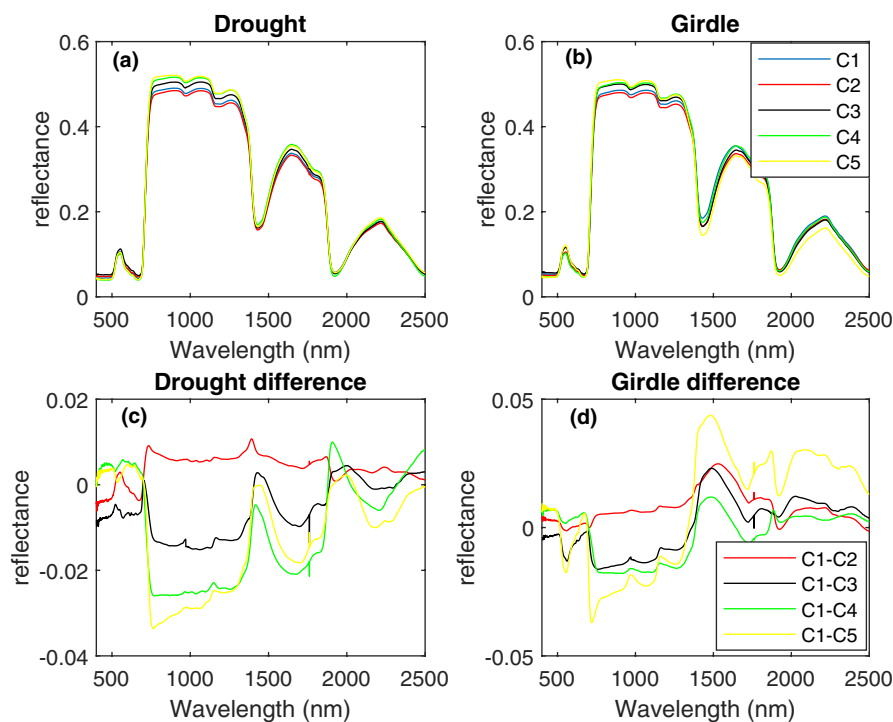
The field campaigns overlapped with the 2016 El Niño in Borneo (Figure 1b). Campaign 1 (C1- Jan-21) took place before the period of peak drought and temperature, C3 (March-16) was conducted during the peak of the drought and high temperatures, and by C5 (June-16)



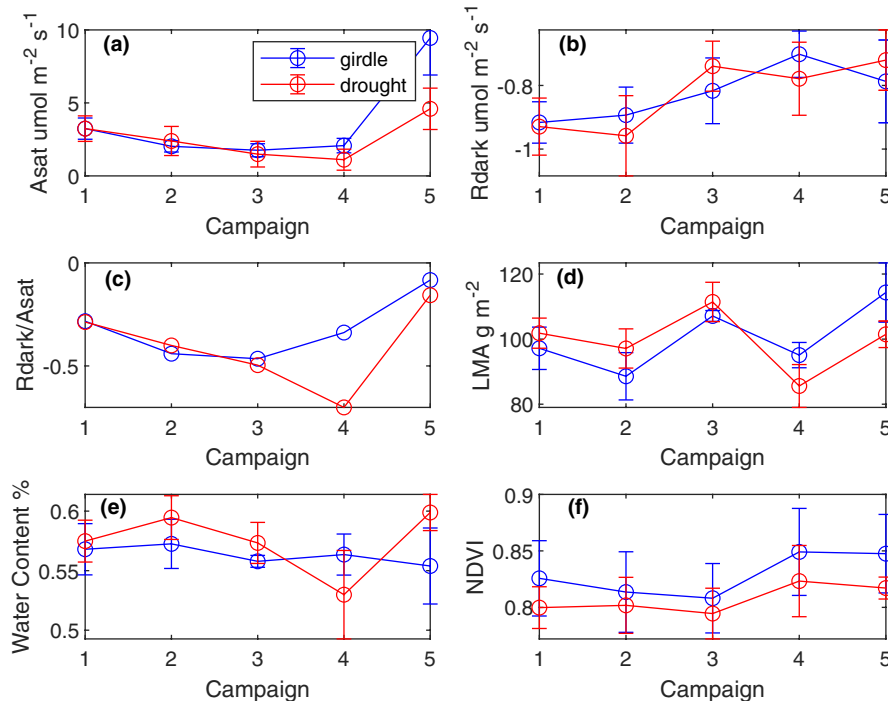
the rains had returned. After C1, all the trees in the girdled plot had their phloem tissue removed in a 10 cm band. Given the downward flux of sugars from the canopy, we might expect an initial build-up of sugars above the girdle followed by eventual tree death as carbon starvation below the girdle impacted tree function, particularly in the roots. Companion papers explore the causes of tree death and the impacts on plant hydraulics and soil respiration (Nottingham, 2020).

There was little change in leaf reflectance (400–2,500 nm) between C1 and C2 (Figure 2) in both the drought and girdled plots. We expected few spectral changes during this short interval between C1 and C2 (Jan-21 to Feb-10) for the natural drought plots, but we were surprised there were also few changes for the girdled plots since these trees experienced a significant trauma. In the later campaigns (C3 to C5 01-Mar to 08-Jun), there were large (~0.03 reflectance units) increases in NIR reflectance (750–1,500 nm) in both the girdled and natural drought plots (Figure 2a,b). Reflectance in the visible wavelengths was lower during peak drought (C3) compared to when the rains returned (C4 and C5). The girdled plots showed a consistent increase in visible reflectance. Spectral reflectance increased in the SWIR bands over time during the drought and there were few changes in the girdled plot except for the final campaign where there was a decrease. Figure 2 displays all spectral data taken during each campaign, and therefore, changes in spectral properties in the girdled plot might also have resulted from species changes because certain tree species died sooner than others, changing the species composition as the experiment continued. To address this, in Figure 5, we compare spectroscopy for only trees that died.

Our average  $A_{\text{sat}}$  values across the campaigns for the girdled plot ( $3.7 \mu\text{mol CO}_2 \text{ m}^{-2} \text{ s}^{-1}$ ) and the drought plot ( $4.7 \mu\text{mol CO}_2 \text{ m}^{-2} \text{ s}^{-1}$ ) were slightly lower, but within 95% confidence intervals of values from a nearby campaign in Borneo (old growth plots -  $4.1 \mu\text{mol CO}_2 \text{ m}^{-2} \text{ s}^{-1}$  (2.7–5.5 = 95% confidence interval) and selectively logged plots -  $7.0 \mu\text{mol CO}_2 \text{ m}^{-2} \text{ s}^{-1}$  (5.7–8.4)) (Both et al., 2019). Our average  $R_{\text{dark}}$  values across the campaigns for the girdled plot ( $-0.82 \mu\text{mol CO}_2 \text{ m}^{-2} \text{ s}^{-1}$ ) and the drought plot ( $-0.83 \mu\text{mol CO}_2 \text{ m}^{-2} \text{ s}^{-1}$ ) were likewise slightly less negative than the values from Both et al., 2019 of  $-1.0 \mu\text{mol CO}_2 \text{ m}^{-2} \text{ s}^{-1}$  ( $-0.9$  to  $-1.2$ ) for the old growth plots and  $-1.3 \mu\text{mol CO}_2 \text{ m}^{-2} \text{ s}^{-1}$  ( $-1.1$  to  $-1.4$ ) for the selectively logged plots. Light-saturated leaf photosynthesis and  $R_{\text{dark}}$  varied between the wet and dry seasons in both plots over the measurement period (Figure 3). Following the return of the rains,  $A_{\text{sat}}$  increased in both the drought and girdled plots in C5. Surprisingly, the surviving girdled trees had the highest photosynthetic rates of all the campaigns in C5 despite the damaged phloem. Dark respiration was at its lowest in C3 and 4 during the peak of the drought. In both groups, changes in  $R_{\text{dark}}$  mirrored those of  $A_{\text{sat}}$ . The ratio  $R_{\text{dark}} / A_{\text{sat}}$  also varied between the wet and dry seasons, with the exception of C4, where the drought plot was less efficient with greater carbon loss per carbon gain. Leaf water content (% leaf water) was also at its lowest in C3 and 4 during the peak of the drought but recovered by C5 (Figure 3e), but we did not find significant effects over time. NDVI was lowest in C3 for both the girdle and drought plots but increased in C4 and C5 (Figure 3f). A repeated measures ANOVA showed no significant differences between  $A_{\text{sat}}$ ,  $R_{\text{d}}$ , and LMA over time between the girdled and drought plots across the



**FIGURE 2** Leaf spectral properties (400–2,500 nm) for the drought (a) and girdled (b) plots for the 5 campaigns (Jan–June 2015). (bottom) The difference (C1–CX, where X = 2–5) in leaf spectral properties for the drought (c) and the girdled (d) plots. In each campaign, we sampled the same trees unless the trees died. Reflectance is reflected incident light between 0–1



**FIGURE 3** Average  $\pm$  se ( $A_{sat}$ ) photosynthetic capacity (a), ( $R_{dark}$ ) leaf dark respiration (b),  $A_{sat}/R_{dark}$  (c) (LMA) leaf mass area (d), % leaf water (e) and NDVI (f) for the 5 campaigns for the control site (red) and the girdled site (blue).  $A_{sat}$  and  $R_d$  were collected at a standard temperature (30°C) during all campaigns. We subtracted the initial difference (2  $\mu\text{mol}/\text{m}^2 \text{ s}^{-1}$ ) in the top panel between the average C1 values to better highlight the impact of the girdling. Peak drought was C3 and the rains returned in C5

four campaigns (C1 was prior to the girdle) suggesting the girdling had little overall impact of on leaf physiology.

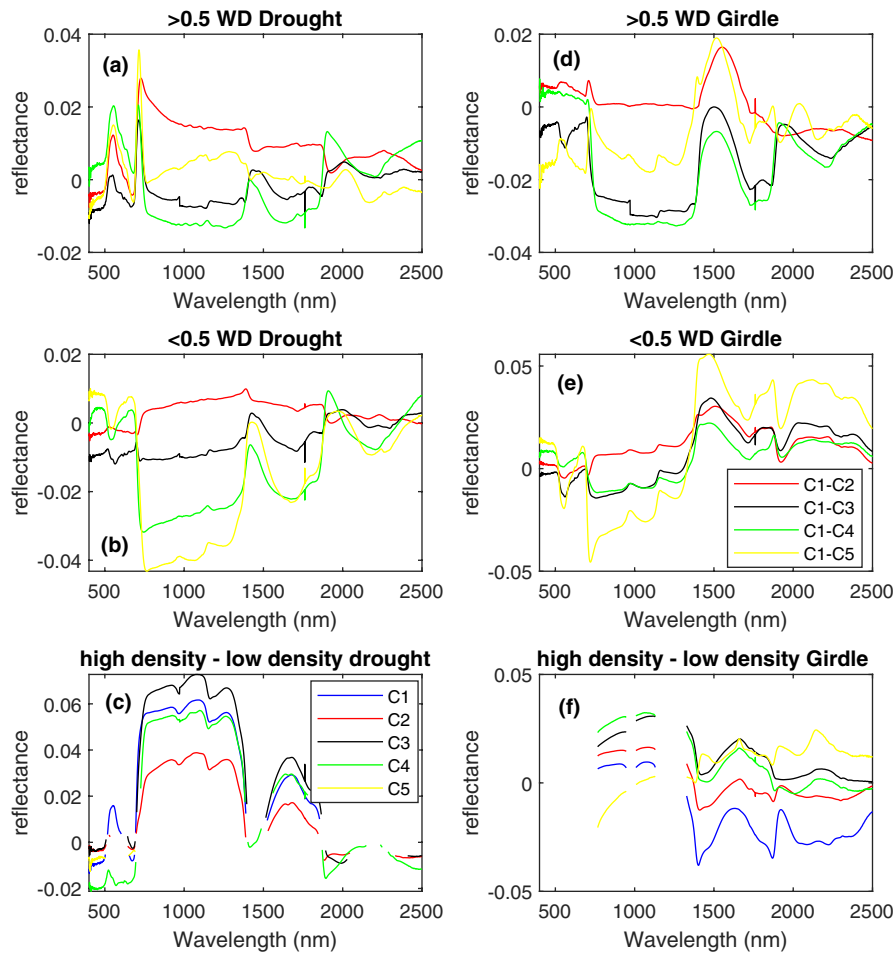
To understand how the drought and girdling impacted leaf spectral properties in different ways and how these link to functional traits, we binned our results into groups of trees with either high ( $>0.5 \text{ g cm}^3 \text{ N} = 359 \text{ leaves/Campaign}$ ) or low wood density ( $<0.5 \text{ g cm}^3 \text{ N} = 830 \text{ leaves/Campaign}$ ) (Figure 4). During the drought, tree species with lower density showed an increase in leaf reflectance compared to species with higher density wood. For example, during the drought, tree species with lower wood density increased leaf reflectance by  $\sim 0.05$  in the NIR and  $\sim 0.01$  SWIR more than tree species with higher density wood, with fewer significant changes ( $p < 0.05$ ) in the visible bands. In contrast, the high wood density tree species show a stronger reaction to the girdling than the low wood density species, again with large increases in reflectance in the NIR and SWIR bands.

We then compared near death leaf reflectance (within 50 days of dying) to leaf reflectance from the same trees, during an earlier period, not near to death (Figure 5). By C5, 38 trees or 18% percent of all girdled trees had died. There were large (0.03–0.05 reflectance units) and significant decreases ( $p < 0.05$ ) in leaf reflectance in the visible bands and the red edge as tree death approached. Close to mortality, there were also large (0.02) and significant increases ( $p < 0.05$ ) in leaf reflectance in NIR and SWIR bands. Next, we investigated how drought conditions, caused by the ENSO event, affected leaf spectral properties in trees which died naturally in the non-girdled control plot. In the control plot, only one tree died from drought that was intensively

sampled for functional traits. We observed similar significant changes along the same pre-death timeline, in leaf reflectance in this tree as observed in the trees that died following the girdling treatment: reductions in reflectance occurred in the red, the NIR and SWIR bands. However, there was a significant peak in the red edge in the opposite direction compared to the girdling study. The wavelengths that show similarities for both types of death were: red (650–700 nm), the NIR (1,000–1,400 nm), and SWIR bands (2000–2,400 nm).

For both the girdled and non-girdled trees, there were highly significant changes ( $p < 0.0001$ ) to the potential carbon balance ( $R_{dark}/A_{sat}$  – Figure 6e,f) of the leaves just prior to death (ie, within 50 days). In both the drought and the girdled plots, there were significant increases in  $R_{dark}$  and significant decreases in  $A_{sat}$  (Figure 6). This combination of increased respiration and decreased photosynthesis should reduce the carbon available to the tree (again dependent on stomatal conductance changes). There was no significant change in LMA among the girdled trees. In contrast, in the tree that died from drought in the non-girdled plot, the leaves had significantly higher LMA and lower % water near to death. We do not know if this was a result of a changing cohort of leaves present on the sampled branch (ie, leaves with lower LMA senesced sooner) or if all leaves changed their LMA via altered density prior to death (less likely as structural carbon is fixed).

Finally, we used PLSR to predict changes in physiology and time to death with spectroscopy (Figure 7). We used the primary weighting (right side of Figure 7) to understand which spectral regions are most important (deviations from 0). Spectroscopy predicted % water and LMA well with an  $r^2$  of 0.72 and 0.74, respectively,



**FIGURE 4** The change in leaf spectral properties (400–2500 nm) between campaigns comparing drought plots for species with high wood density (density > 0.5 g/cm<sup>3</sup> - a), low wood density (density < 0.5 g/cm<sup>3</sup> - b), and the difference (c) through the 5 campaigns. The girdled plots for species with high wood density (density > 0.5 g/cm<sup>3</sup> - d), low wood density (density < 0.5 g/cm<sup>3</sup> - e), and the difference (f). For the difference plots, only significant ( $p < 0.05$ ) spectral regions are shown

and RMSE/mean of 0.07 and 0.14 (similar to many other studies with high precision and high accuracy (Asner & Martin, 2008; Doughty, Asner, et al., 2011) (RMSE = 0.04 and 14.5, RMSE/std = 0.57 and 0.61, # of PLS weights = 30)). The primary weighting is in the NIR and SWIR bands which is typical of traits relating to structure. Spectroscopy predicted maximum photosynthetic rate ( $A_{\text{sat}}$ ) with an  $r^2$  of 0.66 and RMSE/mean of 0.69 (medium precision but low accuracy) (RMSE = 3.3, RMSE/std = 0.74, # of PLS weights = 25/30) and wood density with an  $r^2$  of 0.41 and RMSE/mean of 0.24 (low precision but medium accuracy) (RMSE = 0.12, RMSE/std = 0.94, # of PLS weights = 15). The primary weighting of  $A_{\text{sat}}$  was in the visible bands (likely related to chlorophyll content) and for wood density in NIR and SWIR > 1,000 nm (likely related to variations in LMA and leaf structure). Finally, we predicted time to death with spectroscopy and the PLSR technique with an  $r^2$  of 0.68 and RMSE/mean of 0.55 (medium precision and low accuracy) (RMSE = 82, RMSE/std = 0.81, # of PLS weights = 30). The primary weighting shows similarity with Figure 5 with important spectral regions in the visible (related to photosynthetic characteristics), the NIR (related to structure), and SWIR bands (related to water bands).

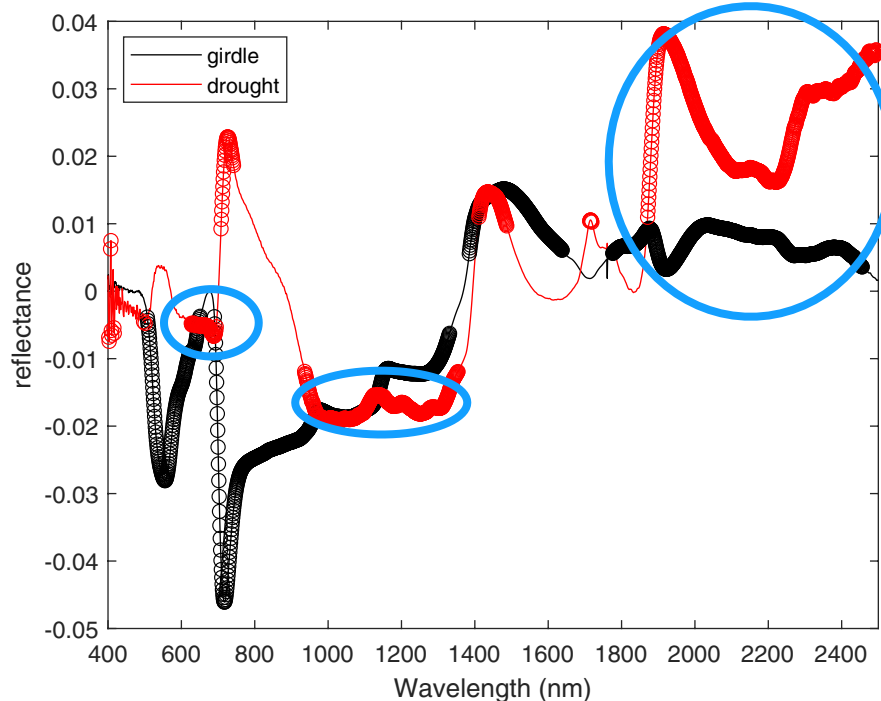
## 4 | DISCUSSION

### 4.1 | Leaf spectroscopy

Identification of tropical trees susceptible to mortality through hyperspectral imagery could provide a powerful tool in examining recently reported increases in tree mortality rates across the tropics (Brienen et al., 2015; Hubau et al., 2020). By contributing to “environmental surveillance,” the use of hyperspectral data would have a wide range of applications from the prediction of tree death from heat stress, pests, pathogens, or illegal logging. Moreover, this technique could enable us to identify potential tipping points in tropical forests, with wider ramifications for the development of adaptive forest management strategies in the future.

Based on these results, future mortality is potentially predictable using hyperspectral data for up to 50 days in advance of tree death (Figure 7). We also observed a tree that died naturally from drought, and saw that there were regions of spectral overlap with the signal from trees killed by girdling in terms of the wavelengths that changed prior to tree death; for example, red (650–700 nm), the NIR





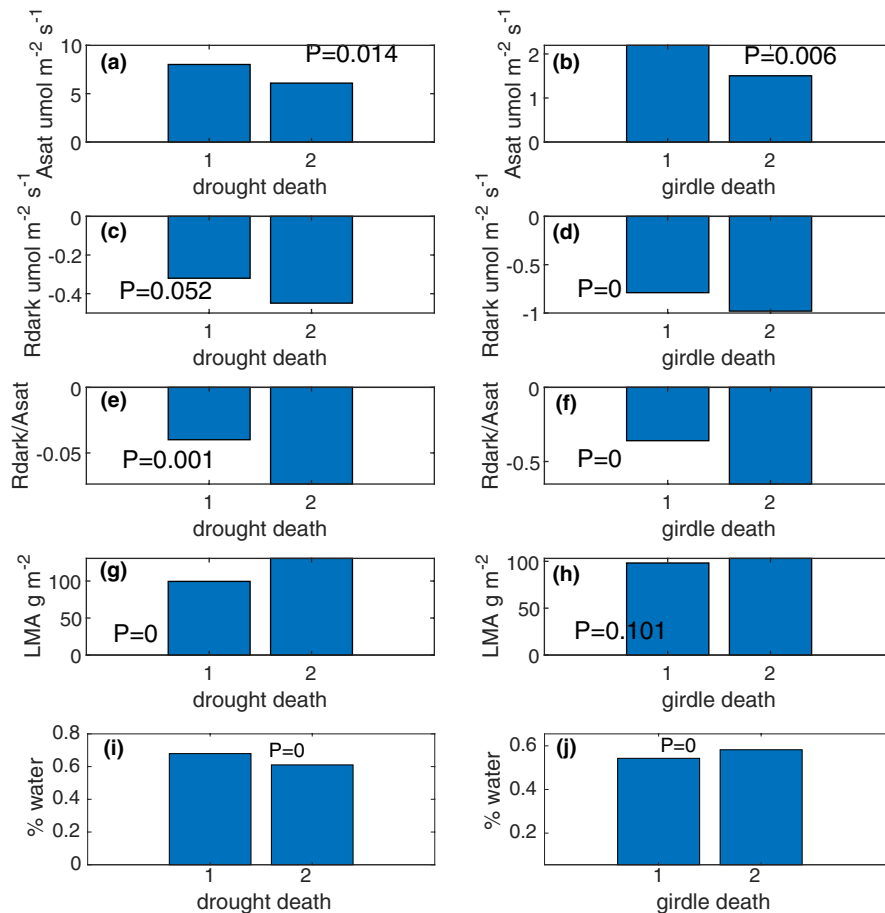
**FIGURE 5** The change (negative is a reduction in reflectance close to death) in leaf spectral properties from healthy leaves (>50 days from death) minus close to death leaves (<50 days from death) on a tree that died of natural drought (red,  $N = 14$  leaves) and trees that died during the girdling experiment (black,  $N = 122$  leaves). Dots show regions of significant change ( $p < 0.05$ ) using a paired  $t$  test. Blue circled areas are key areas of spectral overlap

(1,000–1,400 nm) and SWIR bands (2,000–2,400 nm) (blue circles in Figure 5), but it is difficult to draw conclusions from just one tree. Another venue of remotely sensing stress would be through predicting changes in leaf water content which declined in leaves during the drought (drought and girdled trees) and < 50 days prior to death (only the drought tree) (Figures 3 and 6) and is highly predictable, with high precision and accuracy (Figure 7). This gives us some confidence that the spectral changes may be general to mortality and not specific to girdling-induced mortality. We demonstrate only changes in leaf reflectance and not overall canopy reflectance. It is important to differentiate between leaf versus canopy reflectance (as seen from aircraft or space) because the latter also incorporates forest structural changes (such as variations in LAI, branch architecture, stem density), which were not measured. Leaf spectral properties strongly influence canopy spectral properties especially in certain wavelengths (Asner & Martin, 2008), but changes in other properties, like LAI, would complicate the signal. Leaf-level analyses may also suffer from survivorship bias where the leaves that fare the worst under drought drop first. Large shifts in the spectral regions shown in Figure 5 may be indicative of tree mortality and should be tested with hyperspectral aircraft data in the region for confirmation (Swinfield et al., 2019). A previous study using Hyperion hyperspectral satellite data over an Amazonian drought experiment showed similar declines in magnitude in the NIR and VIS regions as our study (Figure 5) (Asner et al., 2004).

Surprisingly, leaf spectral properties did not vary greatly during the period immediately following tree girdling (~1 week).

Previous studies have quantified changes in non-photosynthetic vegetation to estimate regional selective logging impacts (Asner et al., 2005). Here we show that significant trauma to the trunk (ie, the girdling treatment) did not immediately result in changes to leaf spectral properties, but that leaf spectral properties did change significantly within 50 days of tree death. We hypothesize that > 7 days is the time needed to change the biochemistry, physiology, and metabolism of leaves to respond to substantive environmental stress because we saw little change between C1 and C2. This indicates that > 7 days but < 50 are necessary for leaf spectral changes to occur (Figure 5), which could constrain timing for a potential new technique to identify damage to trees from selective logging.

Do we succeed in predicting mortality because there are changes in short-term physiological status (eg, reduced relative % water content in leaves) or because certain trees are just more likely to die than others due to their constitutive traits (eg, lower LMA linked to a different life history strategy)? In the girdled study, LMA and % water did not change significantly prior to death, but leaf gas exchange metrics did ( $A_{\text{sat}}$  and  $R_{\text{dark}}$ ), shown in the large and significant changes in the visible and red edge bands (Figure 5). However, the drought-associated tree death event was accompanied by a significant change in LMA and % water content, and the spectral analysis showed a further correlation with significant changes in the NIR and water bands (Figure 5). Therefore, it seems a combination of changes in leaf structure, physiological



**FIGURE 6** Comparison of the intensively monitored tree that died during the drought (left) and the girdling experiment (right) for  $A_{\text{sat}}$  (a, b),  $R_{\text{dark}}$  (c, d),  $R_{\text{dark}}/A_{\text{sat}}$  (e, f), LMA (g, h) and % water (i, j) between initial values (1) and values within 50 days of death (2). The P value listed is the level of significance to three digits for a Student's *t* test.  $p = 0$  is a *p* value less than 0.0001

status, and associated reflectance traits combine to enable mortality to be predictable.

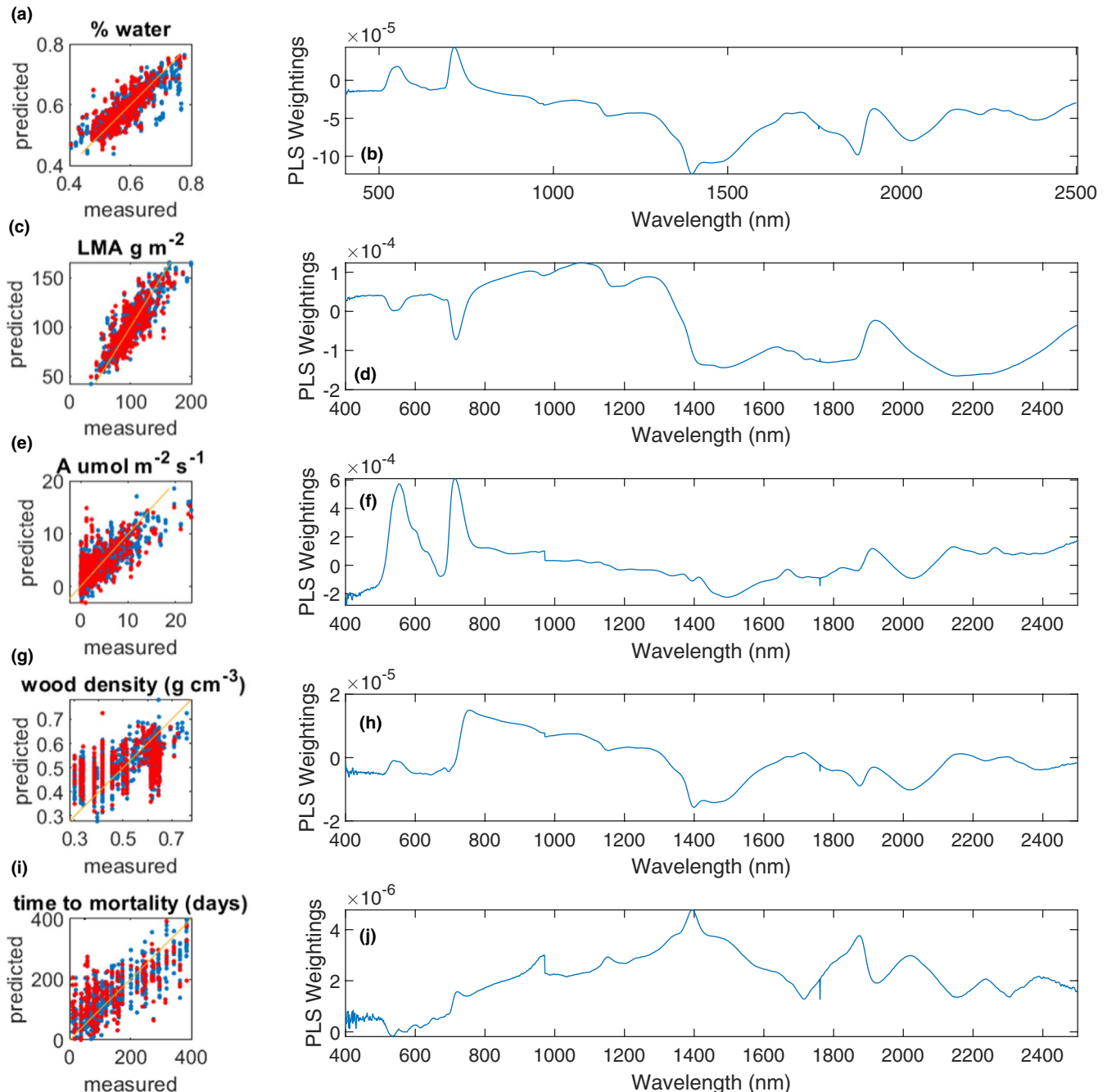
It should be noted that prior to this study our plots had been extensively logged (ie, four times since the 1970s.), with 46 to 54 Mg C ha<sup>-1</sup> cumulative extracted biomass in the area (Riutta et al., 2018). Logging has been shown with hyperspectral imagery in Borneo to lower canopy foliar nutrient concentrations and to decrease nutrient availability (Swinfield et al., 2019). Our results are therefore biased toward logged/low foliar nutrient forests, although our dataset does include late-successional species as well. However, most forests (72%) in the study region have been selectively logged, and our results should be valid for these forests (Bryan et al., 2013).

## 4.2 | Leaf physiology

Leaf dark respiration,  $R_{\text{dark}}$ , was at its lowest (least negative) during the peak of the drought, in campaigns C3 and C4 (Figure 3b). This stands in contrast to some other tropical rainforest leaf respiration studies during natural and artificial drought that have seen

increases in leaf respiration rates (Miranda et al., 2005; Rowland, Lobo-do-Vale, et al., 2015), although recent intensive survey results suggest that the response to experimental drought was taxon-specific rather than observed across a wide range of species (Rowland et al. in review). Leaf  $R_{\text{dark}}$  also did not increase in the leaves of girdled trees despite potential increases in leaf NSC content (as they could not be transported toward the roots following the girdling). Other studies have shown a decrease in overall tree respiration during drought periods as compared to before a drought (Doughty et al., 2015).

We also observed both increased  $R_{\text{dark}}$  and decreased  $A_{\text{sat}}$  50 days prior to tree death (Figure 6), which in combination, are very likely to reduce the carbon available in leaf tissue (although net carbon balance is also dependent on changes in stomatal conductance and light availability). This decreased carbon balance, in turn, could increase the likelihood of carbon starvation (McDowell et al., 2018) and reduce the availability of carbon (or more accurately non-structural carbohydrates) for possible embolism repair in the water conducting xylem tissue (Sala et al., 2012). It is also interesting to note that the highest average photosynthetic capacity ( $A_{\text{sat}}$ ) for the girdled experiment were observed when the



**FIGURE 7** On the left is predictive power (measured versus predicted) for the PLSR analysis with the  $r^2$  and RMSE/mean calculated from the full dataset for various traits including % water (a,  $r^2 = 0.72$ , RMSE = 0.07), LMA (c,  $r^2 = 0.74$ , RMSE/mean = 0.14), Asat (e,  $r^2 = 0.66$ , RMSE/mean = 0.69), wood density (g,  $r^2 = 0.41$ , RMSE/mean = 0.24), and time to tree death (i,  $r^2 = 0.68$ , RMSE/mean = 0.55). Red dots are the data to train the model (70%), and the blue dots are the independent dataset (30%). Sample sizes to train the models are as follows: % water -  $N = 1,035$ , LMA -  $N = 1,028$ , Asat -  $N = 846$ , wood density -  $N = 841$ , tree death -  $N = 543$ . On the right is the primary weighting, which is the PLS weight that explains the most variance in the data, multiplied by variance explained for % water (b), LMA (d), Asat (f), wood density (h), and time to tree death (j)

rains returned. We speculate that might be due to a growth or sink driven response where, after the return of available water there was increased growth (eg, leaf flushing, xylem regrowth) to replace senesced tissue. We hypothesize that the increased growth results in a higher carbon sink leading to a higher demand

for NSC with a consequent increase in  $A_{\text{sat}}$ . Overall, this is strong evidence that photosynthesis remains robust to perturbations, and that growth may be maintained preceding a mortality event as the plant attempts to recover damaged xylem capacity (Rowland, Lobo-do-Vale, et al., 2015; Meir et al., 2015).

## 5 | CONCLUSION

Our key finding is that remote sensing using spectral imagery shows potential to identify trees at imminent risk of death (approximately 50 days prior) with significant ( $p < 0.05$ ) leaf spectral changes in the red (650–700 nm), the NIR (1,000–1,400 nm) and SWIR bands (2,000–2,400 nm). This technique has widespread relevance and applicability for ecological/management surveillance, prediction of future vegetation and forest carbon dynamics. We suggest aircraft campaigns search for a large shift in visible, red edge, and NIR reflectance and compare this to later observed tree mortality or possibly use past data to “hindcast” this technique for validity. For instance, we hypothesize that comparing hyperspectral aircraft flights before and after the 2016 drought might show large shifts in reflectance properties prior to tree mortality (Davies et al., 2019; Swinfield et al., 2019). This could also be of use for hyperspectral satellites like DESIS to predict changes in long-term carbon fluxes associated with tree mortality (Krutz et al., 2019). The large significant changes in leaf reflectance observed here that were shared by both girdling- and drought-killed trees at the same timescale prior to mortality indicate that there could be a spectral indication of tropical tree mortality that has regional or wider application.

## ACKNOWLEDGEMENTS

This study is a product of the BALI consortium, part of the UK Natural Environment Research Council (NERC) Human modified tropical forests program (<http://bali.nerc-hmtf.info/>) and part of the Stability of Altered Forest Ecosystem (SAFE) Project, funded by the Sime Darby Foundation. We would like to acknowledge Dr. Rob Ewers for his role in setting up the SAFE experiment, Elelia Nahun, Dg Ku Shamirah binti Pg Bakar for their contributions to the field campaign, Unding Jami, Ryan Gray, Rostin Jantan, Suhaini Patik and Rohid Kailoh and the BALI and Lombok project research assistants. We also thank the Sabah Biodiversity Centre (SaBC) for permits and SEARRP.

## DATA AVAILABILITY STATEMENT

Data and code that support the findings of this study are available from the Dryad Digital Repository: <https://doi.org/10.5061/dryad.d51c5b01n> (Doughty et al., 2020).

## ORCID

Christopher E. Doughty  <https://orcid.org/0000-0003-3985-7960>

Eleanor R. Thomson  <https://orcid.org/0000-0003-1670-8970>

Alexander Shenkin  <https://orcid.org/0000-0003-2358-9367>

Andrew T. Nottingham  <https://orcid.org/0000-0001-9421-8972>

Walter Huaraca Huasco  <https://orcid.org/0000-0001-5300-4986>

Yadvinder Malhi  <https://orcid.org/0000-0002-3503-4783>

## REFERENCES

- Adams, H. D., Zeppel, M. J. B., Anderegg, W. R. L., Hartmann, H., Landhäuser, S. M., Tissue, D. T., Huxman, T. E., Hudson, P. J., Franz, T. E., Allen, C. D., Anderegg, L. D. L., Barron-Gafford, G. A., Beerling, D. J., Breshears, D. D., Brodribb, T. J., Bugmann, H., Cobb, R. C., Collins, A. D., Dickman, L. T., ... McDowell, N. G. (2017). A multi-species synthesis of physiological mechanisms in drought-induced tree mortality. *Nature Ecology and Evolution*, 1, 1285–1291. <https://doi.org/10.1038/s41559-017-0248-x>
- Anderegg, W. R. L., Klein, T., Bartlett, M., Sack, L., Pellegrini, A. F. A., Choat, B., & Jansen, S. (2016). Meta-analysis reveals that hydraulic traits explain cross-species patterns of drought-induced tree mortality across the globe. *Proceedings of the National Academy of Sciences of the United States of America*, 113, 5024–5029. <https://doi.org/10.1073/pnas.1525678113>
- Anderegg, W. R. L., Konings, A. G., Trugman, A. T., Yu, K., Bowling, D. R., Gabbitas, R., Karp, D. S., Pacala, S., Sperry, J. S., Sulman, B. N., & Zenes, N. (2018). Hydraulic diversity of forests regulates ecosystem resilience during drought. *Nature*, 561, 538–541. <https://doi.org/10.1038/s41586-018-0539-7>
- Asner, G. P., Knapp, D. E., Anderson, C. B., Martin, R. E., & Vaughn, N. (2016). Large-scale climatic and geophysical controls on the leaf economics spectrum. *Proceedings of the National Academy of Sciences of the United States of America*, 113, E4043–E4051. <https://doi.org/10.1073/pnas.1604863113>
- Asner, G. P., Knapp, D. E., Broadbent, E. N., Oliveira, P. J. C., Keller, M., & Silva, J. N. (2005). Ecology: Selective logging in the Brazilian Amazon. *Science*, <https://doi.org/10.1126/science.1118051>
- Asner, G. P., & Martin, R. E. (2008). Spectral and chemical analysis of tropical forests: Scaling from leaf to canopy levels. *Remote Sensing of Environment*, 112, 3958–3970. <https://doi.org/10.1016/j.rse.2008.07.003>
- Asner, G. P., Nepstad, D., Cardinot, G., & Ray, D. (2004). Drought stress and carbon uptake in an Amazon forest measured with spaceborne imaging spectroscopy. *Proceedings of the National Academy of Sciences of the United States of America*, 101, 6039–6044. <https://doi.org/10.1073/pnas.0400168101>
- Bartlett, M. K., Scoffoni, C., & Sack, L. (2012). The determinants of leaf turgor loss point and prediction of drought tolerance of species and biomes: A global meta-analysis. *Ecology Letters*, 15, 393–405. <https://doi.org/10.1111/j.1461-0248.2012.01751.x>
- Bittencourt, P. R. L., Oliveira, R. S., da Costa, A. C. L., Giles, A. L., Coughlin, I., Costa, P. B., Bartholomew, D. C., Ferreira, L. V., Vasconcelos, S. S., Barros, F. V., Junior, J. A. S., Oliveira, A. A. R., Mencuccini, M., Meir, P., & Rowland, L. (2020). Amazonian trees have limited capacity to acclimate plant hydraulic properties in response to long-term drought. *Global Change Biology*, 26, 3569–3584. <https://doi.org/10.1111/gcb.15040>
- Both, S., Riutta, T., Paine, C. E. T., Elias, D. M. O., Cruz, R. S., Jain, A., Johnson, D., Kritzer, U. H., Kuntz, M., Majalap-Lee, N., Mielke, N., Montoya Pilco, M. X., Ostle, N. J., Arn Teh, Y., Malhi, Y., & Burslem, D. F. R. P. (2019). Logging and soil nutrients independently explain plant trait expression in tropical forests. *New Phytologist*, 221, 1853–1865. <https://doi.org/10.1111/nph.15444>
- Brienen, R. J. W., Phillips, O. L., Feldpausch, T. R., Gloor, E., Baker, T. R., Lloyd, J., Lopez-Gonzalez, G., Monteagudo-Mendoza, A., Malhi, Y., Lewis, S. L., Vásquez Martínez, R., Alexiades, M., Álvarez Dávila, E., Alvarez-Loayza, P., Andrade, A., Aragão, L. E. O. C., Araujo-Murakami, A., Arets, E. J. M. M., Arroyo, L., ... Zagt, R. J. (2015). Long-term decline of the Amazon carbon sink. *Nature*, 519, 344–348. <https://doi.org/10.1038/nature14283>
- Bryan, J. E., Shearman, P. L., Asner, G. P., Knapp, D. E., Aoro, G., & Lokes, B. (2013). Extreme differences in forest degradation in Borneo: comparing practices in Sarawak, Sabah, and Brunei. *PLoS ONE*, 8, e69679. <https://doi.org/10.1371/journal.pone.0069679>
- Chave, J., Coomes, D., Jansen, S., Lewis, S. L., Swenson, N. G., & Zanne, A. E. (2009). Towards a worldwide wood economics spectrum. *Ecology Letters*, 12, 351–366. <https://doi.org/10.1111/j.1461-0248.2009.01285.x>
- Clark, D. A. (2004). Sources or sinks? The responses of tropical forests to current and future climate and atmospheric composition. *Philosophical*

- Transactions of the Royal Society of London. Series B: Biological Sciences*, 359, 477–491. <https://doi.org/10.1098/rstb.2003.1426>
- Curran, P. J. (1989). Remote sensing of foliar chemistry. *Remote Sensing of Environment*, 30, 271–278. [https://doi.org/10.1016/0034-4257\(89\)90069-2](https://doi.org/10.1016/0034-4257(89)90069-2)
- da Costa, A. C. L., Galbraith, D., Almeida, S., Portela, B. T. T., da Costa, M., de Athaydes Silva Junior, J., Braga, A. P., de Gonçalves, P. H. L., de Oliveira, A. A. R., Fisher, R., Phillips, O. L., Metcalfe, D. B., Levy, P., & Meir, P. (2010). Effect of 7 yr of experimental drought on vegetation dynamics and biomass storage of an eastern Amazonian rainforest. *New Phytologist*, 187, 579–591. <https://doi.org/10.1111/j.1469-8137.2010.03309.x>
- Davies, A. B., Oram, F., Ancorenaz, M., & Asner, G. P. (2019). Combining behavioural and LiDAR data to reveal relationships between canopy structure and orangutan nest site selection in disturbed forests. *Biological Conservation*, 232, 97–107. <https://doi.org/10.1016/j.biocon.2019.01.032>
- Díaz, S., Kattge, J., Cornelissen, J. H. C., Wright, I. J., Lavorel, S., Dray, S., Reu, B., Kleyer, M., Wirth, C., Colin Prentice, I., Garnier, E., Bönisch, G., Westoby, M., Poorter, H., Reich, P. B., Moles, A. T., Dickie, J., Gillison, A. N., Zanne, A. E., ... Gorné, L. D. (2016). The global spectrum of plant form and function. *Nature*, 529, 167–171.
- Doughty, C. E., Asner, G. P., & Martin, R. E. (2011). Predicting tropical plant physiology from leaf and canopy spectroscopy. *Oecologia*, 165, 289–299. <https://doi.org/10.1007/s00442-010-1800-4>
- Doughty, C. E., Cheesman, A. W., Riutta, T., Thomson, E., Shenkin, A., Nottingham, A. T., Telford, E. M., Huasco, W. H., Majalap, N., Teh, Y. A., Meir, P., & Malhi, Y. (2020). Data from: Predicting tropical tree mortality with leaf spectroscopy. *Dryad Digital Repository*, <https://doi.org/10.5061/dryad.d51c5b01n>
- Doughty, C. E., Field, C. B., & McMillan, A. M. S. (2011). Can crop albedo be increased through the modification of leaf trichomes, and could this cool regional climate? *Climatic Change*, 104(2), 379–387. <https://doi.org/10.1007/s10584-010-9936-0>
- Doughty, C. E., & Goulden, M. L. (2009a). Are tropical forests near a high temperature threshold? *Journal of Geophysical Research: Biogeosciences*, 113(G1). <https://doi.org/10.1029/2007JG000632>
- Doughty, C. E., & Goulden, M. L. (2009b). Seasonal patterns of tropical forest leaf area index and CO<sub>2</sub> exchange. *Journal of Geophysical Research: Biogeosciences*. <https://doi.org/10.1029/2007JG000590>
- Doughty, C. E., Metcalfe, D. B., Girardin, C. A. J., Amézquita, F. F., Cabrera, D. G., Huasco, W. H., Silva-Espejo, J. E., Araujo-Murakami, A., da Costa, M. C., Rocha, W., Feldpausch, T. R., Mendoza, A. L. M., da Costa, A. C. L., Meir, P., Phillips, O. L., & Malhi, Y. (2015). Drought impact on forest carbon dynamics and fluxes in Amazonia. *Nature*, 519, 78–82. <https://doi.org/10.1038/nature14213>
- Doughty, C. E., Santos-Andrade, P. E., Goldsmith, G. R., Blonder, B., Shenkin, A., Bentley, L. P., Chavana-Bryant, C., Huaraca-Huasco, W., Díaz, S., Salinas, N., Enquist, B. J., Martin, R., Asner, G. P., & Malhi, Y. (2017). Can leaf spectroscopy predict leaf and forest traits along a peruvian tropical forest elevation gradient? *Journal of Geophysical Research: Biogeosciences*, 122, 2952–2965. <https://doi.org/10.1002/2017JG003883>
- Ewers, R. M., Didham, R. K., Fahrig, L., Ferraz, G., Hector, A., Holt, R. D., Kapos, V., Reynolds, G., Sinun, W., Snaddon, J. L., & Turner, E. C. (2011). A large-scale forest fragmentation experiment: the Stability of Altered Forest Ecosystems Project. *Philosophical Transactions of the Royal Society B: Biological Sciences*, 366, 3292–3302. <https://doi.org/10.1098/rstb.2011.0049>
- Fyllas, N. M., Quesada, C. A., & Lloyd, J. (2012). Deriving Plant Functional Types for Amazonian forests for use in vegetation dynamics models. *Perspectives in Plant Ecology, Evolution and Systematics*, 14, 97–110. <https://doi.org/10.1016/j.ppees.2011.11.001>
- Geladi, P., & Kowalski, B. R. (1986). Partial least-squares regression: A tutorial. *Analytica Chimica Acta*, 2670, 1–17. [https://doi.org/10.1016/0003-2670\(86\)80028-9](https://doi.org/10.1016/0003-2670(86)80028-9)
- Gvozdevaite, A., Oliveras, I., Domingues, T. F., Peprah, T., Boakye, M., Afriyie, L., da Silva Peixoto, K., de Farias, J., Almeida de Oliveira, E., Almeida Farias, C. C., dos Santos Prestes, N. C. C., Neyret, M., Moore, S., Schwantes Marimon, B., Marimon Junior, B. H., Adu-Bredu, S., & Malhi, Y. (2018). Leaf-level photosynthetic capacity dynamics in relation to soil and foliar nutrients along forest–savanna boundaries in Ghana and Brazil. *Tree Physiology*, 38, 1912–1925. <https://doi.org/10.1093/treephys/tpy117>
- Hubau, W., Lewis, S. L., Phillips, O. L., Affum-Baffoe, K., Beeckman, H., Cuní-Sánchez, A., Daniels, A. K., Ewango, C. E. N., Fauset, S., Mukinzi, J. M., Sheil, D., Sonké, B., Sullivan, M. J. P., Sunderland, T. C. H., Taedoum, H., Thomas, S. C., White, L. J. T., Abernethy, K. A., Adu-Bredu, S., ... Zemagho, L. (2020). Asynchronous carbon sink saturation in African and Amazonian tropical forests. *Nature*, 579, 80–87. <https://doi.org/10.1038/s41586-020-2035-0>
- Jacquemoud, S., Verhoef, W., Baret, F., Bacour, C., Zarco-Tejada, P. J., Asner, G. P., François, C., & Ustin, S. L. (2009). PROSPECT + SAIL models: A review of use for vegetation characterization. *Remote Sensing of Environment*, 113, S56–S66. <https://doi.org/10.1016/j.rse.2008.01.026>
- Krutz, D., Müller, R., Knodt, U., Günther, B., Walter, I., Sebastian, I., Säuberlich, T., Reulke, R., Carmona, E., Eckardt, A., & Venus, H. (2019). The Instrument Design of the DLR Earth Sensing Imaging Spectrometer (DESI). *Sensors*, 19, 1622. <https://doi.org/10.3390/s19071622>
- Malhi, Y., Gardner, T. A., Goldsmith, G. R., Silman, M. R., & Zelazowski, P. (2014). Tropical Forests in the Anthropocene. *Annual Review of Environment and Resources*, 39, 125–159. <https://doi.org/10.1146/annurev-environ-030713-155141>
- Maréchaux, I., Bartlett, M. K., Sack, L., Baraloto, C., Engel, J., Joetzier, E., & Chave, J. (2015). Drought tolerance as predicted by leaf water potential at turgor loss point varies strongly across species within an Amazonian forest. *Functional Ecology*, 29, 1268–1277. <https://doi.org/10.1111/1365-2435.12452>
- McDowell, N., Allen, C. D., Anderson-Teixeira, K., Brando, P., Brienens, R., Chambers, J., Christoffersen, B., Davies, S., Doughty, C., Duque, A., Espirito-Santo, F., Fisher, R., Fontes, C. G., Galbraith, D., Goodsman, D., Grossiord, C., Hartmann, H., Holm, J., Johnson, D. J., ... Xu, X. (2018). Drivers and mechanisms of tree mortality in moist tropical forests. *New Phytologist*, 219, 851–869. <https://doi.org/10.1111/nph.15027>
- Meir, P., M. Mencuccini, O. Binks, A. C. L. da Costa, L. Ferreira, and L. Rowland. (2018). Short-term effects of drought on tropical forest do not fully predict impacts of repeated or long-term drought: gas exchange versus growth. *Philosophical Transactions of the Royal Society B*. <https://doi.org/10.1098/rstb.2017.0311>
- Meir, P., Wood, T. E., Galbraith, D. R., Brando, P. M., Da Costa, A. C. L., Rowland, L., & Ferreira, L. V. (2015). Threshold Responses to Soil Moisture Deficit by Trees and Soil in Tropical Rain Forests: Insights from Field Experiments. *BioScience*, 65, 882–892. <https://doi.org/10.1093/biosci/biv107>
- Miranda, E. J., Vourlitis, G. L., Filho, N. P., Priante, P. C., Campelo, J. H., Suli, G. S., Fritzen, C. L., de Almeida Lobo, F., & Shiraiwa, S. (2005). Seasonal variation in the leaf gas exchange of tropical forest trees in the rain forest–savanna transition of the southern Amazon Basin. *Journal of Tropical Ecology*, 21, 451–460. <https://doi.org/10.1017/S0266467405002427>
- Morton, D. C., Nagol, J., Carabjal, C. C., Rosette, J., Palace, M., Cook, B. D., Vermote, E. F., Harding, D. J., & North, P. R. J. (2014). Amazon forests maintain consistent canopy structure and greenness during the dry season. *Nature*, 506, 221–224. <https://doi.org/10.1038/nature13006>
- Nepstad, D. C., Tohver, I. M., David, R., Moutinho, P., & Cardinot, G. (2007). Mortality of large trees and lianas following experimental drought in an amazon forest. *Ecology*, 88, 2259–2269. <https://doi.org/10.1890/06-1046.1>



- Niinemets, Ü. (2001). Global-scale climatic controls of leaf dry mass per area, density, and thickness in trees and shrubs. *Ecology*, 82, 453–469. [10.1890/0012-9658\(2001\)082\[0453:GSCCOL\]2.0.CO;2](https://doi.org/10.1890/0012-9658(2001)082[0453:GSCCOL]2.0.CO;2)
- Nottingham, A. (2020). Large Contribution of Recent Photosynthate to Soil Respiration in Dipterocarpaceae-dominated Tropical Forest Revealed by Girdling. *EcoEvoRxiv*, <https://doi.org/10.32942/osf.io/df6zg>
- Nunes, M. H., Both, S., Bongalov, B., Brelsford, C., Khoury, S., Burslem, D. F. R. P., Philipson, C., Majalap, N., Riutta, T., Coomes, D. A., & Cutler, M. E. (2019). Changes in leaf functional traits of rainforest canopy trees associated with an El Niño event in Borneo. *Environmental Research Letters*, 14, 85005. <https://doi.org/10.1088/1748-9326/ab2eae>
- Phillips, O. L., Aragao, L. E. O. C., Lewis, S. L., Fisher, J. B., Lloyd, J., Lopez-Gonzalez, G., Malhi, Y., Monteagudo, A., Peacock, J., Quesada, C. A., van der Heijden, G., Almeida, S., Amaral, I., Arroyo, L., Aymard, G., Baker, T. R., Banki, O., Blanc, L., Bonal, D., ... Torres-Lezama, A. (2009). Drought sensitivity of the amazon rainforest. *Science*, 323, 1344–1347. <https://doi.org/10.1126/science.1164033>
- Poorter, H., Niinemets, Ü., Poorter, L., Wright, I. J., & Villar, R. (2009). Causes and consequences of variation in leaf mass per area (LMA): A meta-analysis. *New Phytologist*, 182, 565–588. <https://doi.org/10.1111/j.1469-8137.2009.02830.x>
- Poorter, L., Wright, S. J., Paz, H., Ackerly, D. D., Condit, R., Ibarra-Manríquez, G., Harms, K. E., Licona, J. C., Martínez-Ramos, M., Mazer, S. J., Muller-Landau, H. c., Peña-Claros, M., Webb, C. O., & Wright, I. J. (2008). Are functional traits good predictors of demographic rates? evidence from five neotropical forests. *Ecology*, 89, 1908–1920. <https://doi.org/10.1890/07-0207.1>
- Rifai, S. W., Girardin, C. A. J., Berenguer, E., del Aguila-Pasquel, J., Dahlsjö, C. A. L., Doughty, C. E., Jeffery, K. J., Moore, S., Oliveras, I., Riutta, T., Rowland, L. M., Murakami, A. A., Addo-Danso, S. D., Brando, P., Burton, C., Ondo, F. E., Duah-Gyamfi, A., Amézquita, F. F., Freitag, R., ... Malhi, Y. (2018). ENSO Drives interannual variation of forest woody growth across the tropics. *Philosophical Transactions of the Royal Society B: Biological Sciences*, 373, 20170410. <https://doi.org/10.1098/rstb.2017.0410>
- Rifai, S. W., Li, S., & Malhi, Y. (2019). Coupling of El Niño events and long-term warming leads to pervasive climate extremes in the terrestrial tropics. *Environmental Research Letters*, 14, 105002. <https://doi.org/10.1088/1748-9326/ab402f>
- Riutta, T., Malhi, Y., Kho, L. K., Marthews, T. R., Huaraca Huasco, W., Khoo, M. S., Tan, S., Turner, E., Reynolds, G., Both, S., Burslem, D. F. R. P., Teh, Y. A., Vairappan, C. S., Majalap, N., & Ewers, R. M. (2018). Logging disturbance shifts net primary productivity and its allocation in Bornean tropical forests. *Global Change Biology*, 24(7), 2913–2928. <https://doi.org/10.1111/gcb.14068>
- Rowland, L., da Costa, A. C. L., Galbraith, D. R., Oliveira, R. S., Binks, O. J., Oliveira, A. A. R., Pullen, A. M., Doughty, C. E., Metcalfe, D. B., Vasconcelos, S. S., Ferreira, L. V., Malhi, Y., Grace, J., Mencuccini, M., & Meir, P. (2015). Death from drought in tropical forests is triggered by hydraulics not carbon starvation. *Nature*, 528, 119–122. <https://doi.org/10.1038/nature15539>
- Rowland, L., Lobo-do-Vale, R. L., Christoffersen, B. O., Melém, E. A., Kruijt, B., Vasconcelos, S. S., Domingues, T., Binks, O. J., Oliveira, A. A. R., Metcalfe, D., Costa, A. C. L., Mencuccini, M., & Meir, P. (2015). After more than a decade of soil moisture deficit, tropical rainforest trees maintain photosynthetic capacity, despite increased leaf respiration. *Global Change Biology*, 21, 4662–4672. <https://doi.org/10.1111/gcb.13035>
- Rowland, L., Zaragoza-Castells, J., Bloomfield, K. J., Turnbull, M. H., Bonal, D., Burban, B., Salinas, N., Cosío, E., Metcalfe, D. J., Ford, A., Phillips, O. L., Atkin, O. K., & Meir, P. (2017). Scaling leaf respiration with nitrogen and phosphorus in tropical forests across two continents. *New Phytologist*, 214, 1064–1077. <https://doi.org/10.1111/nph.13992>
- Sala, A., Woodruff, D. R., & Meinzer, F. C. (2012). Carbon dynamics in trees: Feast or famine? *Tree Physiology*, <https://doi.org/10.1093/treephys/tpz143>
- Saleska, S. R., Didan, K., Huete, A. R., & Da Rocha, H. R. (2007). Amazon forests green-up during 2005 drought. *Science*, 318, 612. <https://doi.org/10.1126/science.1146663>
- Samanta, A., Ganguly, S., Hashimoto, H., Devadiga, S., Vermote, E., Knyazikhin, Y., Nemani, R. R., & Myneni, R. B. (2010). Amazon forests did not green-up during the 2005 drought. *Geophysical Research Letters*, 37. <https://doi.org/10.1029/2009GL042154>
- Sapes, G., Roskilly, B., Dobrowski, S., Maneta, M., Anderegg, W. R. L., Martinez-Vilalta, J., & Sala, A. (2019). Plant water content integrates hydraulics and carbon depletion to predict drought-induced seedling mortality. *Tree Physiology*, 39, 1300–1312. <https://doi.org/10.1093/treephys/tpz062>
- Serbin, S. P., Singh, A., McNeil, B. E., Kingdon, C. C., & Townsend, P. A. (2014). Spectroscopic determination of leaf morphological and biochemical traits for northern temperate and boreal tree species. *Ecological Applications*, 24, 1651–1669. <https://doi.org/10.1890/13-2110.1>
- Sevanto, S., McDowell, N. G., Dickman, L. T., Pangle, R., & Pockman, W. T. (2014). How do trees die? A test of the hydraulic failure and carbon starvation hypotheses. *Plant, Cell & Environment*, 37, 153–161. <https://doi.org/10.1111/pce.12141>
- Swinfield, T., Both, S., Riutta, T., Bongalov, B., Elias, D., Majalap-Lee, N., Ostle, N., Svátek, M., Kvasnica, J., Milodowski, D., Jucker, T., Ewers, R. M., Zhang, Y. I., Johnson, D., Teh, Y. A., Burslem, D. F. R. P., Malhi, Y., & Coomes, D. (2019). Imaging spectroscopy reveals the effects of topography and logging on the leaf chemistry of tropical forest canopy trees. *Global Change Biology*, 26, 989–1002. <https://doi.org/10.1111/gcb.14903>
- Thirumalai, K., DInezio, P. N., Okumura, Y., & Deser, C. (2017). Extreme temperatures in Southeast Asia caused by El Niño and worsened by global warming. *Nature Communications*, 8, 1–8. <https://doi.org/10.1038/ncomms15531>
- Ustin, S. L., Asner, G. P., Gamon, J. A., Fred Huemmrich, K., Jacquemoud, S., Schaepman, M., & Zarco-Tejada, P. (2006). Retrieval of quantitative and qualitative information about plant pigment systems from high resolution spectroscopy. *International Geoscience and Remote Sensing Symposium (IGARSS)*. <https://doi.org/10.1109/IGARSS.2006.517>
- Ustin, S. L., Gitelson, A. A., Jacquemoud, S., Schaepman, M., Asner, G. P., Gamon, J. A., & Zarco-Tejada, P. (2009). Retrieval of foliar information about plant pigment systems from high resolution spectroscopy. *Remote Sensing of Environment*, 113, S67–S77. <https://doi.org/10.1016/j.rse.2008.10.019>
- Walsh, R. P. D., & Newbery, D. M. (1999). The ecoclimatology of Danum, Sabah, in the context of the world's rainforest regions, with particular reference to dry periods and their impact. *Philosophical Transactions of the Royal Society of London. Series B: Biological Sciences*, 354, 1869–1883. <https://doi.org/10.1098/rstb.1999.0528>
- Wright, I. J., Reich, P. B., Westoby, M., Ackerly, D. D., Baruch, Z., Bongers, F., Cavender-Bares, J., Chapin, T., Cornelissen, J. H. C., Diemer, M., Flexas, J., Garnier, E., Groom, P. K., Gulias, J., Hikosaka, K., Lamont, B. B., Lee, T., Lee, W., Lusk, C., ... Villar, R. (2004). The worldwide leaf economics spectrum. *Nature*, 428, 821–827. <https://doi.org/10.1038/nature02403>
- Wright, S. J., Kitajima, K., Kraft, N. J. B., Reich, P. B., Wright, I. J., Bunker, D. E., Condit, R., Dalling, J. W., Davies, S. J., Díaz, S., Engelbrecht, B. M. J., Harms, K. E., Hubbell, S. P., Marks, C. O., Ruiz-Jaen, M. C., Salvador, C. M., & Zanne, A. E. (2010). Functional traits and the

- growth-mortality trade-off in tropical trees. *Ecology*, 91, 3664–3674. <https://doi.org/10.1890/09-2335.1>
- Wu, J., Kobayashi, H., Stark, S. C., Meng, R., Guan, K., Tran, N. N., Gao, S., Yang, W., Restrepo-Coupe, N., Miura, T., Oliveira, R. C., Rogers, A., Dye, D. G., Nelson, B. W., Serbin, S. P., Huete, A. R., & Saleska, S. R. (2018). Biological processes dominate seasonality of remotely sensed canopy greenness in an Amazon evergreen forest. *New Phytologist*, 217, 1507–1520. <https://doi.org/10.1111/nph.14939>
- Zanne, A. E., Westoby, M., Falster, D. S., Ackerly, D. D., Loarie, S. R., Arnold, S. E. J., & Coomes, D. A. (2010). Angiosperm wood structure: Global patterns in vessel anatomy and their relation to wood density

and potential conductivity. *American Journal of Botany*, 97, 207–215. <https://doi.org/10.3732/ajb.0900178>

**How to cite this article:** Doughty CE, Cheesman AW, Riutta T, et al. Predicting tropical tree mortality with leaf spectroscopy. *Biotropica*. 2021;53:581–595. <https://doi.org/10.1111/btp.12901>